

The Kinetics of Ethanol Fermentation Based on Adsorption Processes

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Currently, almost all ethanol fermentation kinetic models are based on the Monod equation. Such models belong to empirical models, hence are not suitable for the optimization and expansion of real processes. In this work, a new ethanol fermentation kinetic model, in which all parameters had definite physical meaning, was established based on the adsorption mechanism. Through batch fermentation experiment with different initial glucose concentrations and comparison with existing model, this model has the advantage of high precision and few parameters. It also possessed obvious regulation in the model parameter that, with the increase in initial glucose concentration, the maximum specific growth rate μ_{\max} and adsorption equilibrium constant K_s of glucose decreased and adsorption equilibrium constant K_p of ethanol increased. This model provides a new perspective for kinetics research of fermentation.

Key words: *Fermentation, biofuel, kinetics, biochemical engineering*

Introduction

Currently, the oil crisis and serious environmental problems related to fossil fuels have caused the world to turn more attention toward biofuels production, especially ethanol fermentation from biomass. An appropriate kinetic model of ethanol fermentation would be a powerful instrument for fermentation efficiency increase and process optimization. The well-known Monod equation has been widely applied and modified to describe different fermentation processes.^{1–7} Unlike the Michaelis-Menten equation describing the mechanism of enzyme catalysis, the Monod equation is purely empirical with no mechanistic basis and hardly meets functional requirements to support large-scale production, process scale-up and optimization. In this paper, a simple kinetic model of ethanol fermentation is proposed, which provides a new perspective for kinetics research of fermentation.

Model development

Cell growth, substrate consumption, and product formation were taken into account in the developed kinetic model of ethanol fermentation. Most fundamental observations concerning cell growth were used to describe the behaviour of the given microbial system. The following first-order kinetic equation of cell growth was widely used:

$$r_x = \frac{d\gamma_x}{dt} = \mu \gamma_x, \quad (1)$$

where r_x denotes cell growth rate and γ_x is cell concentration. The ideal expression of specific growth rate μ has been dominated by an empirical model originally proposed by Monod:

$$\mu = \frac{\mu_{\max} \gamma_s}{k_s + \gamma_s}, \quad (2)$$

where k_s is half-saturation coefficient and γ_s is glucose concentration. The microorganism can be also considered as a catalyst. Kinetic equation based on the catalysis theory in chemical engineering is generally in the form

$$r_x = K \gamma_x \theta_s, \quad (3)$$

where K is the rate constant, and θ_s is the fractional cell surface coverage with adsorbed glucose.

In the case of ethanol production by yeast cells, three essential steps were involved, including transportation of glucose into cells, glucose metabolism, as well as ethanol formation and passing through the membrane. Since glucose does not freely permeate biological membrane, its cellular uptake requires transporters to drive it across the membrane. The rate of sugar uptake in yeast cells is controlled by the glucose transportation.⁸ The facilitated transport of glucose moving into the cells is mediated by the reversible binding of glucose to the transport protein. This involves some kind of conformational change within the protein and neither substrates consumption nor chemical bonds making or breaking were found in this reversible process. The specific binding of glucose to the transporters can be considered as adsorption and

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θ_s is assumed as the fraction of transporter on the surface of membrane to which glucose is adsorbed.

Owing to limitations of the classical Monod equation, an additional product inhibition term is present in general which is responsible for the delay in growth caused by the ethanol accumulation. r_x is given by *Lun et al.*:⁹

$$r_x = \frac{\mu_{\max} \gamma_S}{k_S + \gamma_S} \frac{k_P}{k_P + \gamma_P} \left(1 - \frac{\gamma_P}{\gamma_{P,\max}}\right) \gamma_X, \quad (4)$$

where k_P refers to the product inhibition constant, and $\gamma_{P,\max}$ is the product concentration above which cells do not grow.

The exact mechanism of ethanol inhibition is yet to be fully understood. The reported proposal for the inhibition effects may result from the denaturation of glycolytic enzymes, damaging and altering membrane organizations, and inhibition of glucose, maltose, amino acid and ammonium transport, etc.¹⁰

The rate of glucose transport depends on the amount of the carrier available and the activity of transporters.¹¹ The stability of transport protein is easily affected by the addition of ethanol, which is a protein conformation destabilizer, through the increment of protein secondary structure, disruption of protein native structure, and inducement of amyloid formation.¹² The transport can be blocked by ethanol due to the denaturation of protein through the formation of competitive bonds between the alcohol and the non-polar residues of the protein, and results in the inhibition of glucose fermentation. Thereby, the transporters in the yeast cell membrane can be divided into three parts: transporters occupied by glucose (θ_s), transporters occupied by ethanol (θ_p), and free transporters (θ_v). The value of θ_s can be calculated according to the Langmuir adsorption model.

It can be assumed that the adsorption of glucose (G) and ethanol (E) to the transporters (σ) occurred simultaneously.



The adsorption rate varied directly with substance concentration and the fraction of free transporters on the surface of membrane ($\theta_v = 1 - \theta_s - \theta_p$). The desorption rate of glucose or ethanol is directly related to the value of θ_s or θ_p . Consequently, the glucose adsorption rate (r_{aS}) and desorption rate (r_{dS}) can be represented as follows:

$$r_{aS} = k_{aS} \gamma_S (1 - \theta_s - \theta_p), \quad (6)$$

$$r_{dS} = k_{dS} \theta_s, \quad (7)$$

where k_{aS} is the adsorption constant of glucose, k_{dS} is the desorption constant of glucose.

The adsorption rate (r_{aP}) and desorption rate (r_{dP}) of ethanol could be expressed by:

$$r_{aP} = k_{aP} \gamma_P (1 - \theta_s - \theta_p), \quad (8)$$

$$r_{dP} = k_{dP} \theta_p, \quad (9)$$

where k_{aP} and k_{dP} are the adsorption constant and desorption constant of ethanol, respectively.

When adsorption of glucose or ethanol reached equilibrium, the net adsorption rate is zero.

$$r_{aS} = r_{dS} \quad (10)$$

$$r_{aP} = r_{dP} \quad (11)$$

Combining Eqs. 6–11 gives:

$$\theta_s = \frac{K_S \gamma_S}{1 + K_S \gamma_S + K_P \gamma_P}, \quad (12)$$

$$\theta_p = \frac{K_P \gamma_P}{1 + K_S \gamma_S + K_P \gamma_P}, \quad (13)$$

where K_S is the adsorption equilibrium constant of glucose and was expressed as $K_S = k_{aS}/k_{dS}$. K_P is the adsorption equilibrium constant of ethanol and was expressed as $K_P = k_{aP}/k_{dP}$.

The complete kinetic equation accounting for ethanol inhibition is obtained by merging Eq. 12 and Eq. 3.

$$r_x = k \frac{K_S \gamma_S}{1 + K_S \gamma_S + K_P \gamma_P} \gamma_X \quad (14)$$

Here k can be designated as μ_{\max} which leads to the common form of kinetic equation.

$$r_x = \mu_{\max} \frac{K_S \gamma_S}{1 + K_S \gamma_S + K_P \gamma_P} \gamma_X. \quad (15)$$

This equation appears totally different from the usual kinetic equations for cell cultures due to the simultaneous presence of γ_S and γ_P in the denominator.

If the impact of ethanol on the transporter is omitted, Eq. 14 becomes

$$r_x = k \frac{K_S \gamma_S}{1 + K_S \gamma_S} \gamma_X = \frac{k \gamma_S}{\frac{1}{K_S} + \gamma_S} \gamma_X. \quad (16)$$

It is in accordance with the Monod equation, and indicates that the kinetic pattern of microorganisms can be explained by the adsorption mechanism as well.

For verification, the proposed model was applied to batch

ethanol fermentation by yeast subjected to simulate the kinetic pattern and compared with the fermentation profiles and prediction of the classic model as Eq. 4.

Since the product of ethanol fermentation is related to cell growth, the ethanol production rate and substrate consumption rate are given by Lee *et al.*¹³

$$r_p = \frac{r_x}{Y_{X/P}} \quad (17)$$

$$r_s = \frac{r_x}{Y_{X/S}}, \quad (18)$$

where, r_s and r_p are glucose consumption rate and ethanol formation rate, respectively. $Y_{X/S}$ is yield coefficient of biomass on glucose and $Y_{X/P}$ is yield coefficient of biomass on ethanol.

Experiments

Commercial ethanol instant active dry yeast (*Saccharomyces cerevisiae*) was used in this study, which was purchased from Angle Yeast Co., Ltd. Experiments were carried out to study the kinetics of batch fermentation of yeast at 35 °C in a Biotech-5BG fermentor (Shanghai Baoxin Bioengineering Equipment CO., LTD.), which was agitated by a magnetic stirrer at 200 rpm and controlled at pH 5.0. For preparation of yeast inoculum, 1 g dry yeast was added to 20 ml sterile water containing 2 % glucose, and incubated at 37 °C for 1 h. The inoculum concentration was 1 g l⁻¹. The experiments were conducted with various initial glucose concentrations of 110 g l⁻¹, 160 g l⁻¹, and 220 g l⁻¹ respectively. The culture medium was composed of 3 g l⁻¹ yeast extraction, 5 g l⁻¹ KH₂PO₄, 2 g l⁻¹ (NH₄)₂SO₄, 0.2 g l⁻¹ MgSO₄, and 0.1 g l⁻¹ CaCl₂. All the reagents used in this study were of AR grade. All experimental data were the average of duplicate results, with variation smaller than ± 5 %.

Ethanol concentration was determined by HP4890D gas chromatography. Glucose concentration was measured by the dinitrosalicylic (DNS) colorimetric method. Biomass concentration was measured as optical density at 640 nm by interpolation from a pre-established dry weight calibration curve.

Results and discussion

Parameters in the kinetic model were estimated by nonlinear least-squares regression of the experimental data using lsqnonlin function in Matlab. Figs. 1–3 represent the simulations for cell growth, glucose consumption, and ethanol production during batch culture carried out by the classic model (Eq. 4) and the proposed model (Eq. 15). The required parameters in kinetic equations were calculated and are listed in Table 1. Simulated results indicated that, when the initial concentrations of glucose were 110 g l⁻¹ and 160 g l⁻¹, both models could satisfactorily simulate the fermentation pattern. Furthermore, the values of parameters $Y_{X/P}$ and $Y_{X/S}$ predicted from the two models are very close to each other. When the initial concentration of glucose reached 220 g l⁻¹, the difference between the experimental data and the pre-

dicted value increased and turned out to be significant. It is probably due to the high substrate inhibition which was not included in both models. But considering that such high level of glucose is too high for practical purposes, the proposed model is much more useful for the optimization of ethanol batch fermentations by yeast compared with the classic model, because of its lower value of calculated residues, higher accuracy of prediction, and less parameters involved.

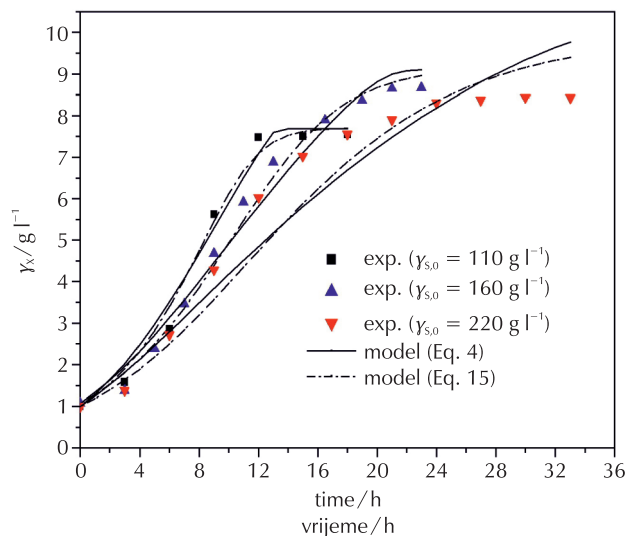


Fig. 1 – Experimental results and simulation of cell concentration at different initial concentrations of glucose (110 g l⁻¹, 160 g l⁻¹, and 220 g l⁻¹)

Slika 1 – Eksperimentalni rezultati i modeliranje koncentracije stanica pri različitim početnim koncentracijama glukoze (110 g l⁻¹, 160 g l⁻¹ i 220 g l⁻¹)

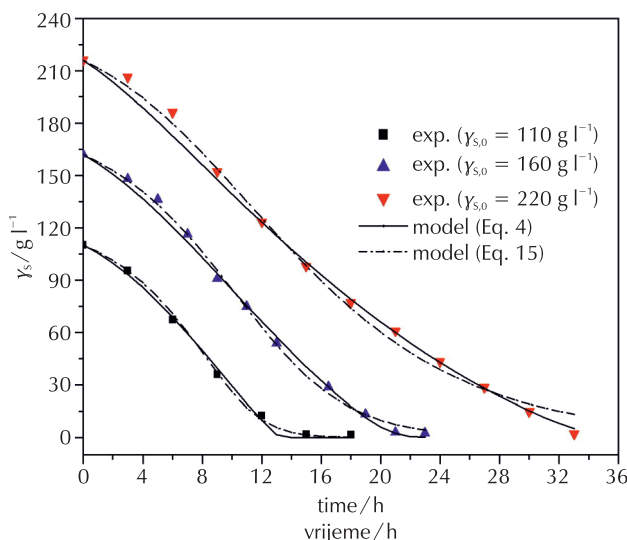


Fig. 2 – Experimental results and simulation of glucose concentration at different initial concentrations of glucose (110 g l⁻¹, 160 g l⁻¹, and 220 g l⁻¹)

Slika 2 – Eksperimentalni rezultati i modeliranje koncentracije glukoze pri različitim početnim koncentracijama glukoze (110 g l⁻¹, 160 g l⁻¹ i 220 g l⁻¹)

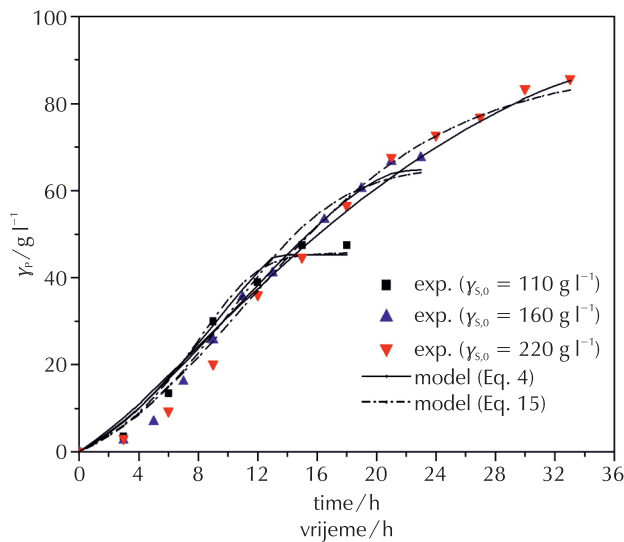


Fig. 3 – Experimental results and simulation of ethanol concentration at different initial concentrations of glucose (110 g l^{-1} , 160 g l^{-1} , and 220 g l^{-1})

Slika 3 – Eksperimentalni rezultati i modeliranje koncentracije etanola pri različitim početnim koncentracijama glukoze (110 g l^{-1} , 160 g l^{-1} i 220 g l^{-1})

Table 1 – Kinetics parameters

Tablica 1 – Kinetički parametri

	$Y_{S,0} = 110 \text{ g l}^{-1}$	$Y_{S,0} = 160 \text{ g l}^{-1}$	$Y_{S,0} = 220 \text{ g l}^{-1}$	
Model A (Eq. 15)	μ_{\max}/h^{-1}	0.2174	0.1790	0.1704
	$K_S/\text{g l}^{-1}$	19.6015	13.2139	6.5788
	$K_P/\text{g l}^{-1}$	16.7864	21.7157	23.7452
	$Y_{X/P}$	0.1467	0.1229	0.1011
	$Y_{X/S}$	0.0608	0.0499	0.0414
	res.*	53.44	159.45	400.9077
Model B (Eq. 4)	μ_{\max}/h^{-1}	0.2631	0.2301	0.2342
	$k_S/\text{g l}^{-1}$	1.1683	4.6143	2.5230
	$k_P/\text{g l}^{-1}$	40.5013	39.2565	28.1250
	$Y_{P,\max}/\text{g l}^{-1}$	129.7881	128.8183	121.8918
	$Y_{X/P}$	0.1477	0.1239	0.1027
	$Y_{X/S}$	0.0606	0.0496	0.0415
res.*	74.4205	220.2505	516.2401	

$$* \text{ residue} = \sum_{i=1}^n (C_{\text{exp}} - C_{\text{calc}})^2$$

From Table 1, it is obvious that there is no general relationship between the initial glucose concentrations and the values of μ_{\max} and k_S of model B, due to the fact that parameters in this empirical model represent no meaningful aspects of reality. It affects the capability of this model to provide a reliable basis for optimization and further scale-up of ethanol batch fermentations process by yeast. However, in model A, the predicted values of μ_{\max} decreased with an increase in the initial glucose concentration, indicating substrate inhibition on cell growth. The values of K_S , however, decreased with the increase in the initial glucose concentration suggesting that the increment of osmotic pressure of the solution caused by the high glucose concentration had a negative effect on the activity of transport protein in yeast cells, which resulted in an increase in the value of rate constant for glucose desorp-

tion from transport protein (r_{ds}). Furthermore, the ethanol molecule is smaller than glucose and more easily adsorbed by the transport protein, therefore the increment value of K_S will lead to a decrement in K_P due to the competition between glucose and ethanol. This phenomenon is enhanced by the increase in ethanol concentration.

Conclusion

The proposed kinetic model of batch ethanol fermentation by yeast based on the irreversible adsorption mechanism could bring a revolution in the transfer of the empirical model to a physical model for fermentation kinetics, since every parameter in this model provides physical meaningful aspects of reality. Compared with the classic model, the proposed model is more practicable for the optimization of ethanol fermentation due to its higher accuracy of prediction, and less parameters involved. Furthermore, the variation in the kinetic parameters with the initial glucose concentration is obtained and represents as follows. The predicted values of μ_{\max} and K_S decreased with an increase in the initial glucose concentration, while an increment in K_P was observed.

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List of symbols and abbreviations Popis simbola i kratica

- K_P – adsorption equilibrium constant of ethanol, g l^{-1}
– adsorpcijska ravnotežna konstanta etanola, g l^{-1}
- K_S – adsorption equilibrium constant of glucose, g l^{-1}
– adsorpcijska ravnotežna konstanta glukoze, g l^{-1}
- k – reaction constant, h^{-1}
– reakcijska konstanta, h^{-1}
- k_{aP} – ethanol adsorption constant, h^{-1}
– adsorpcijska konstanta etanola, h^{-1}
- k_{aS} – glucose adsorption constant, h^{-1}
– adsorpcijska konstanta glukoze, h^{-1}
- k_{dP} – ethanol desorption constant, $\text{g l}^{-1} \text{h}^{-1}$
– desorpcijska konstanta etanola, $\text{g l}^{-1} \text{h}^{-1}$
- k_{dS} – glucose desorption constant, $\text{g l}^{-1} \text{h}^{-1}$
– desorpcijska konstanta glukoze, $\text{g l}^{-1} \text{h}^{-1}$
- k_P – ethanol inhibition constant, g l^{-1}
– inhibicijska konstanta etanola, g l^{-1}
- k_S – half-saturation coefficient, g l^{-1}
– koeficijent poluzasićenosti, g l^{-1}
- r_{aP} – ethanol adsorption rate, $\text{g l}^{-1} \text{h}^{-1}$
– brzina adsorpcije etanola, $\text{g l}^{-1} \text{h}^{-1}$
- r_{aS} – glucose adsorption rate, $\text{g l}^{-1} \text{h}^{-1}$
– brzina adsorpcije glukoze, $\text{g l}^{-1} \text{h}^{-1}$
- r_{dP} – ethanol desorption rate, $\text{g l}^{-1} \text{h}^{-1}$
– brzina desorpcije etanola, $\text{g l}^{-1} \text{h}^{-1}$
- r_{dS} – glucose desorption rate, $\text{g l}^{-1} \text{h}^{-1}$
– brzina desorpcije glukoze, $\text{g l}^{-1} \text{h}^{-1}$
- r_P – ethanol formation rate, $\text{g l}^{-1} \text{h}^{-1}$
– brzina stvaranja etanola, $\text{g l}^{-1} \text{h}^{-1}$

r_s	– glucose consumption rate, $g\ l^{-1}\ h^{-1}$ – brzina potrošnje glukoze, $g\ l^{-1}\ h^{-1}$
r_x	– cell growth rate, $g\ l^{-1}\ h^{-1}$ – brzina rasta stanica, $g\ l^{-1}\ h^{-1}$
$Y_{X/S}$	– yield coefficient of biomass on glucose – koeficijent prinosa biomase na glukozu
$Y_{X/P}$	– yield coefficient of biomass on ethanol – koeficijent prinosa biomase na etanolu
Y_P	– ethanol concentration, $g\ l^{-1}$ – koncentracija etanola, $g\ l^{-1}$
Y_S	– glucose concentration, $g\ l^{-1}$ – koncentracija glukoze, $g\ l^{-1}$
$Y_{S,0}$	– initial glucose concentration, $g\ l^{-1}$ – početna koncentracija glukoze, $g\ l^{-1}$
Y_X	– cell concentration, $g\ l^{-1}$ – koncentracija stanica, $g\ l^{-1}$
θ_p	– fractional cell surface coverage with adsorbed glucose – pokrivenost površine stanica adsorbiranom glukozom
θ_s	– fractional cell surface coverage with adsorbed ethanol – pokrivenost površine stanica adsorbiranim etanolom
θ_v	– fractional free cell surface – udjel slobodne površine stanica
μ	– specific growth rate, h^{-1} – specifična brzina rasta, h^{-1}
DNS	– dinitrosalicylic acid – dinitrosalicilna kiselina
E	– ethanol – etanol
G	– glucose – glukoza
max	– maximum – maksimum
rpm	– revolutions per minute – broj okretaja u minuti
σ	– transporter – transporter

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SAŽETAK**Kinetika fermentacije etanola temeljena na procesima adsorpcije***Z. Liu* i X. Li*

Trenutačno se gotovo svi kinetički modeli fermentacije etanola temelje na Monodovoj jednadžbi. Takvi modeli se ubrajaju u iskustvene modele, pa nisu prikladni za optimizaciju i uvećanje stvarnih procesa. U ovom je radu razvijen novi kinetički model fermentacije etanola na temelju mehanizma adsorpcije, u kojem svi parametri imaju određeni fizikalni smisao. Šaržnom fermentacijom s različitim početnim koncentracijama glukoze i usporedbom s postojećim modelom utvrđeno je da model ima prednost u visokoj preciznosti i manje parametara te se jednostavno regulira jer se maksimalna specifična brzina rasta μ_{\max} i adsorpcijska ravnotežna konstanta glukoze K_s smanjuju s povećanjem početne koncentracije glukoze, a povećava adsorpcijska ravnotežna konstanta etanola K_p . Spomenuti model pruža novu točku gledišta za istraživanje kinetike fermentacije.

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